

“Sebocytes’ makeup” - Novel mechanisms and concepts in the physiology of the human sebaceous glands

Balázs I. Tóth · Attila Oláh · Attila G. Szöllösi · Gabriella Czifra · Tamás Bíró

Received: 30 November 2010 / Revised: 8 February 2011 / Accepted: 11 February 2011 / Published online: 8 March 2011
© Springer-Verlag 2011

Abstract The pilosebaceous unit of the human skin consists of the hair follicle and the sebaceous gland. Within this “mini-organ”, the sebaceous gland has been neglected by the researchers of the field for several decades. Actually, it was labeled as a reminiscence of human development (“a living fossil with a past but no future”), and was thought to solely act as a producer of sebum, a lipid-enriched oily substance which protects our skin (and hence the body) against various insults. However, due to emerging research activities of the past two decades, it has now become evident that the sebaceous gland is not only a “passive” cutaneous “relic” to establish the physico-chemical barrier function of the skin against constant environmental challenges, but it rather functions as an “active” neuro-immuno-endocrine cutaneous organ. This review summarizes recent findings of sebaceous gland research by mainly focusing on newly discovered physiological functions, novel regulatory mechanisms, key events in the pathology of the gland, and future directions in both experimental and clinical dermatology.

Keywords Human sebaceous gland · Lipid synthesis · Immuno-endocrine functions · Signal transduction · Acne vulgaris

Preface

The sebaceous gland (glandula sebacea) is a holocrine gland located in the dermis of the skin, where it is primarily

associated with hair follicles forming the pilosebaceous unit [111]. Its cells are the sebocytes, and their main, well-known function is the production of sebum (tallow) [147]. Sebaceous glands can be found in the skin of all terrestrial mammals, but the density of the glands in the different regions of the body, as well as the composition of the sebum, exhibit a marked species specificity. The primary isolated sebaceous glands and sebocytes cannot be kept in culture for a long time because of their rapid and excessive differentiation [36, 106]. Taking in account that most of the pathological alterations of the sebaceous gland functions (e.g., acne vulgaris) are exclusively human diseases, the identification of relevant models has been a critical question of human sebaceous gland research. In the last decade, the *in vitro* animal models, e.g., preputial gland of rodents [103, 153] or sebaceous gland of hamster ear [99], were mainly substituted by human sebaceous gland-derived cell lines, such as the SZ95 [171], Seb-1 [144], and Seb-E6E7 [66]. These models have provided a new insight into human sebaceous gland biology and have broadened our understanding of the molecular mechanisms and regulation of different sebaceous functions [169, 172]. This review summarizes recent findings of sebaceous gland research by mainly focusing on newly discovered functions (e.g., immunological functions) and novel regulatory mechanisms.

Diverse functions of an ancient gland

Development, differentiation, and regeneration

Differentiation of the pilosebaceous unit occurs in the embryonic stage between months 2 and 4 of gestation. During this process, a complex, in many aspects still unresolved, signaling interplay between dermal mesenchy-

B. I. Tóth · A. Oláh · A. G. Szöllösi · G. Czifra · T. Bíró (✉)
Department of Physiology, Medical and Health Science Center,
Research Center for Molecular Medicine, University of Debrecen,
4032 Debrecen, Nagyerdei krt. 98, PO Box 22, Hungary
e-mail: biro@phys.dote.hu

mal cells and the embryonic epidermis induces the formation of the dermal papilla of the hair follicle, which initiates the final differentiation of the pilosebaceous unit. The source of the sebocytes is the basal layer of the epidermis and the progenitor cells are identical to the cells which form the outer root sheath of the hair follicle [26]. During embryonic life, the sebaceous glands may contribute to the formation of vernix caseosa [26, 165].

Recent findings shed a light on the molecular mechanisms of the sebaceous gland differentiation. The differentiation of pilosebaceous stem cells depends on Sox9 signaling since this signal is important for both hair follicle and sebaceous gland differentiation [84]. However, during the following steps, the development of the two structures is regulated differentially. The lineage choice of the progenitors can be regulated by the Wnt target β -catenin; i.e., the presence of β -catenin promotes hair follicle differentiation whereas its inhibition, e.g., by Smad7 overexpression, shifts the process towards the sebaceous lineage. The further proliferation of sebocyte precursors is stimulated by Indian Hedgehog signaling [4, 45, 83] while the bone morphogenic protein pathway, similar to Wnt signaling, was reported to inhibit sebaceous differentiation [42].

During embryonic development, a population of unipotent sebaceous stem cells is established which express the B lymphocyte-induced maturation protein 1 (Blimp1) transcription factor [48]. These cells appear to be essential in the renewing of sebocytes (and hence regenerating the mini-organ) in the adult skin; however, stem cells from the bulge region of hair follicle are also capable of forming sebaceous glands [110, 111]. In addition, Blimp1 inhibits the expression of c-myc which suppression is essential in normal sebaceous gland homeostasis; consequently, deletion of Blimp1 results an overexpression of c-myc and causes the hyperplasia of sebaceous glands [48]. Other results show that sebaceous cell lines can differentiate into both sebocytes and keratinocytes, which suggests the presence of bi-potential progenitor stem cells among them [66].

The adult, fully developed sebaceous gland can be divided into three zones which contain cells at different stages of differentiation. The *peripheral zone* is composed of small, mitotically active cells. During their differentiation, these cells move towards the center of the gland, lose their mitotic activity, increase their size, and accumulate lipid droplets, forming the *maturation zone*. In the *central necrosis zone*, the terminally differentiated sebocytes disintegrate and release their content via holocrine secretion [110, 147]. This continuous differentiation program is under the control of various paracrine, endocrine, and neural mediators acting on a wide array of receptors expressed by sebocytes [162]. Some of the newly recognized factors, which influence/regulate the physiological

processes of the sebaceous gland, will be discussed in detail below.

“Classical” functions of the sebaceous glands—sebum production: the barrier and beyond

Sebum is mainly composed of neutral lipids, with a relatively high amount of triglycerides, free fatty acids, wax esters, cholesterol, and squalene. Among these, squalene and wax esters are unique and typical components of the sebum [97, 104, 136]. The secreted sebum covers the fur and the surface of the skin, and forms the majority of skin surface lipids. In addition, a much smaller lipid fraction is produced by epidermal keratinocytes, which mostly fill the intercellular spaces between keratinocytes and ensure the skin permeability barrier [93].

In animals, sebum plays important roles in the impregnation of fur and thermal insulation, while in some species the sebaceous glands have specialized to produce pheromones. Since these functions are mostly unrecognizable in humans, it has been a long-standing view that the human sebaceous gland is an evolutionary relic [100]. However, the unique composition of the sebaceous lipids plays important roles in the skin barrier function. For instance, waxes can be more resistant to oxidation than other lipids and can improve the water resistance of the surface [93]. Others reported the potential role of squalene and its peroxides in the process of protection against sunburn and UV radiation [88]. It was also found that sebum contains the antioxidant vitamin E [146]. Furthermore, sebaceous lipids are not only a part of the physico-chemical barrier of the skin, but they form a biological barrier as well; indeed, sebum contains various lipids with antimicrobial activity (see below).

Besides the well-documented barrier function, other roles are also ascribed to human sebum. One of them is the supposed role in thermoregulation. Under cold circumstances, sebum can form a water-repellent layer whereas in warmer environments, it is transformed to a more fluidic form. The latter can serve as an emulsifier for eccrine sweat by decreasing its surface tension and thereby helping to keep the sweat on the skin surface which increases the efficacy of evaporation [68, 102]. In addition, sebum may influence not only the eccrine, but also the apocrine sweat functions. This new theory hypothesizes that sebum, produced by axillary sebaceous glands, can serve as vehicle for fragrances, and, as such, it may influence odor and can also play a role in the interpersonal communication [165].

Immunological functions of the sebaceous glands

Recent findings also show that functions of sebocytes may go far beyond the production of sebum and the formation of the passive cutaneous barrier. Via numerous paracrine,

endocrine, and immunological mechanisms, sebaceous glands greatly contribute to the physiological homeostatic function of the skin.

Sebocytes participate in the regulation of immunological functions and inflammatory processes. They are capable of producing different (mostly pro-inflammatory) cytokines and lipid-derived inflammatory mediators. Among cytokines, the expression of interleukin (IL) 1 α , IL-1 β , IL-6, IL-8/CXCL-8, and tumor necrosis factor- α (TNF α ; but not IL-10 and IL-12) was reported in human sebaceous glands and cultured sebocytes [3, 81]. It was also shown that the expression and release of cytokines could be affected by various factors, such as, e.g., inflammatory signals. While the presence of *Propionibacterium acnes* up-regulated the expression of TNF α and IL-8/CXCL-8, bacterial lipopolysaccharide (LPS), treatment elevated the levels of IL-1 α as well [81]. Likewise, arachidonic acid (AA) and the Ca²⁺ ionophore A23187 increased the release of IL-6 and IL-8 [3] whereas IL-1 β was found to stimulate the release of IL-8 [170]. Interestingly, the activation of the Ca²⁺ permeable transient receptor potential vanilloid-1 (TRPV1) channel did not affect the release of IL-6, but it did decrease that of IL-1 β [149]. Various hormones and neuropeptides are also able to influence the cytokine release of sebocytes; these include hypothalamic and pituitary hormones, i.e., corticotropin-releasing hormone (CRH) and α -melanocyte stimulating hormone (α MSH), and the neuropeptide substance P (SP); hence, these mediators may affect the inflammatory processes of these cells (see also below).

As inflammatory signals, certain lipid mediators may also play a crucial role. These substances are derivatives of AA produced by the cyclooxygenase (COX) or lipoxygenase (LOX) pathways [37, 139]. The sebocytes express key enzymes of both pathways—i.e., 5-LOX and leukotriene A₄-hydrolase (LTA₄-hydrolase) of the LOX pathway and COX-1 and 2—and are able to synthesize leukotriene B₄ (LTB₄) and prostaglandin E₂ (PGE₂), which production can be enhanced by inflammatory stimuli, for example UV irradiation or AA administration [3, 160].

The above listed wide array of inflammatory mediators produced by the sebaceous gland may act as key players in the pathogenesis of inflammatory syndromes such as, e.g., acne vulgaris. The development of acne requires multiple pathological processes such as comedo-genesis due to the hyperproliferation of keratinocytes, increased lipid synthesis of sebocytes with alterations in the lipid content of the sebum, and the proliferation of pathogenic microorganisms such as *P. acnes*. In parallel, the production of inflammatory mediators (produced both by keratinocytes and sebocytes) is increased which, in turn, attract the “on-site” invasion of (first) CD4⁺ T lymphocytes then neutrophil granulocytes to infiltrate the affected pilosebaceous unit. Of further importance, novel etiological models emphasize that

acne may develop without the colonization of pathogenic microorganisms, provided that some other factors (e.g., enhanced effect of androgenic hormones, activation of peroxisome proliferator-activated receptors (PPAR), SP-mediated stress response, other hormonal effects, etc.) increase the production of the inflammatory mediators and induce hyperseborrhea [33, 59, 161, 168].

Sebocytes of the sebaceous glands can be considered as not only “producers” of potential pathogenic factors but these cells are also part of the innate immune system. As we mentioned above, sebum contains lipids with antimicrobial activity. These lipids are especially effective against Gram-positive bacteria, such as *Staphylococcus aureus* (including methacillin-resistant strains), *Streptococcus salivarius*, *Fusobacterium nucleatum*, *Pseudomonas aeruginosa*, *Escherichia coli*, and *P. acnes* [32, 154]. Chemically, these antimicrobial lipids can be both saturated (e.g., lauric acid, C_{12:0}) and unsaturated (e.g., sapienic acid, C_{16:1 Δ 6}) fatty acids. Importantly, saturated fatty acids with shorter chain length form monomers in aqueous solutions with higher probability. Since the antimicrobial activity of these lipids is attributed chiefly to the monomer forms, the differences seen in their chemical natures suggest that shorter lipids exhibit greater antimicrobial activities than long-chain fatty acids. Furthermore, antimicrobial fatty acids, such as, e.g., the monounsaturated fatty acid (MUFA) sapienic acid, may be synthesized by sebocytes in the form of triglyceride esters and then can be liberated by bacterial triglyceride hydrolysis or by epidermal acid lipase [32]. The antimicrobial role of MUFAs was further supported by the flake homozygote mouse model. These mice exhibit an impaired stearoylcoenzyme A desaturase 1 enzyme function and hence are unable to produce the MUFAs palmitoleate (C_{16:1}) and oleate (C_{18:1}). Consequently, these animals show signs of severe dermatitis, and are more vulnerable to Gram-positive bacterial infections [39].

Finally, it should be emphasized that, similar to the release of pro-inflammatory immune mediators, sebocytes can also secrete certain peptides/proteins such as, e.g., antimicrobial peptides. Indeed, it was shown on SZ95 sebocytes that these cells express functional cathelicidin [63], β -defensins [81], and histone H4 [62], and that the expression of these peptides can be induced by bacterial stimuli. In addition, sebocytes also express Toll-like receptors [81, 86, 87, 162] further supporting the concept that sebaceous glands are indeed indisputable players in innate immunity.

Extended hormonal control of the sebaceous gland

Our concept about the skin has been dramatically revised in the past 20 years. The function of our largest organ is no

more restricted for the physico-chemical barrier, thermo-regulatory and sensory mechanisms; rather, the skin is introduced as a complex endocrine organ of the human body since it is a source as well as a target of a plethora of (neuro)endocrine hormones and autocrine/paracrine mediators [116, 127, 129, 163]. As an integral part of the human skin, the pilosebaceous unit and its sebocytes also play an active role in these endocrine functions [12, 21].

Effects of steroid hormones

It has been known for a long time that the sebum production of sebaceous glands is stimulated by androgens [100, 147] (Table 1). The presence of androgen receptors was found both in situ on human sebaceous glands [22, 95] and in vitro on human sebocytes [35]. Moreover, testosterone and 5 α -dihydrotestosterone (5 α -DHT) was shown to increase proliferation of sebocytes in vitro [35, 171]. Using primary isolated sebocytes, it was also reported that the location of the sebaceous gland influences the effect of androgens; namely, these hormones were more effective in increasing the proliferation of facial sebocytes than on non-facial ones [1, 164]. Intriguingly, androgen hormones alone failed to modulate lipid synthesis of cultured sebocytes [35, 171]. However, in the presence of certain co-activators such as, e.g., linoleic acid which stimulates PPARs [107, 108], androgens may exert lipogenic actions.

Of great importance, sebocytes seem to be much more than simple “passive targets” of the effects of androgens. Recent evidence suggests that they are also capable of metabolizing and synthesizing androgen hormones and hence play a central role in cutaneous androgen homeostasis [19, 167]. These cells express members of the P450 side-chain cleavage system which converts cholesterol to pregnenolone [144]. Sebocytes also express the androgen metabolizing enzymes 3 β -hydroxysteroid dehydrogenase/ Δ 5-4-isomerase, 17 β -hydroxysteroid dehydrogenase, 5 α -reductase-1, and 3 β -hydroxysteroid dehydrogenase. Intriguingly, sebocytes are reportedly able to synthesize testosterone and also to convert testosterone into the more effective 5 α -DHT which process, like lipid synthesis, was promoted by a simultaneous activation of PPARs [73]. Furthermore, these cells can inactivate testosterone by converting it to androstenedione and further to 5 α -androstenedione [35, 113].

In contrast to the actions of androgens, estrogens were originally described to suppress the lipid production of the sebaceous gland [26, 44]. However, recent reports have provided conflicting data; namely, although expressions of estrogen receptors α and β and progesterone receptor were shown on sebaceous glands [95], the female sexual steroids 17 β -estradiol and progesterone were found to influence neither proliferation nor lipid synthesis of SZ95 sebocytes in vitro [72].

Less data were reported about the sebaceous effects of corticosteroid hormones. The presence of the enzymatic apparatus needed for the production of corticosteroids as well as the corticosteroids (such as, e.g., cortisol) themselves were extensively documented in the skin [52, 121, 122, 131–135]. These facts postulate that the sebaceous gland may also be a target of the locally produced corticosteroids. Early observations suggested that prednisolone failed to induce sebum synthesis but glucocorticoids might have a permissive effect on the sebaceous gland activity [101]. Topically administered glucocorticoids were found to decrease human sebum production [65]. In addition, corticosteroids were shown to be able to stimulate proliferation [172] and inhibit lipid synthesis of cultured sebocytes [18] which findings suggested that corticosteroids may exert a differentiation-inhibiting effect.

Finally, another steroid hormone, the active vitamin D₃ metabolite calcitriol deserves mentioning. The 1,25-dihydroxy vitamin D₃ is produced locally by the keratinocytes of the skin and can regulate the differentiation of several cutaneous cell types [5, 6, 46]. Importantly, SZ95 sebocytes express the 1 α , 24, and 25-hydroxylases and vitamin D receptor. Furthermore, calcitriol was shown to influence the lipid content, regulate proliferation, reduce the IL-6 and IL-8 release, and up-regulate cathelicidin antimicrobial peptide expression on SZ95 sebocytes [57, 63]. Hence, calcitriol may also function as a key regulator of sebocyte biology.

Effects of growth hormones and growth factors

Sebaceous gland functions are also under the control of growth-promoting hormones and growth factors (Table 1). The “oily skin” is a characteristic for acromegaly (a syndrome that develops as a result of overproduction of growth hormone, GH) and, moreover, the identification of GH receptors in the sebaceous gland in situ [67, 85] suggested the potential role of GH, mRNA of which was identified also in human skin [123], in sebaceous gland functions. Indeed, in a rat prepubertal sebocyte model, GH accelerated the differentiation of sebocytes in vitro but did not significantly influence proliferation. In contrast, insulin-like growth factor-I (IGF-I), which was shown to mediate several physiological actions of GH in various tissues, had a minor effect on differentiation but it markedly increased the proliferation of sebocytes. Intriguingly, the “universal growth hormone” insulin was found to stimulate both proliferation and differentiation, and it augmented the effects of GH, IGF-I, and 5 α -DHT [25]. In human sebaceous cell lines, both GH and IGF-I were able to enhance lipid synthesis, IGF-I being the more effective one [72]. Of further importance, the effect of IGF-I was found to be mediated by the PI-3-kinase/Akt/sterol response element-binding protein-1 (SREBP1) pathway [137, 138].

Table 1 Mediators and agents influencing the functions of sebocytes

Agents	Potential targets	Possible effects
Bacterial stimuli (e.g., presence of <i>P. acnes</i> , LPS)	Toll-like receptor-2, 4, and 6 (TLR-2, TLR-4, TLR-6) [81, 86, 87]	β -defensins \uparrow , cathelicidin \uparrow , tumor necrosis factor- α (TNF α) \uparrow , interleukin-8 (IL-8) \uparrow , IL-1 α \uparrow [62, 63, 81]
AA	Unknown (protein kinases?, peroxisome proliferator-activated receptors—PPARs?)	IL-6 \uparrow , IL-8 \uparrow , leukotriene B ₄ (LTB ₄) \uparrow , lipid synthesis \uparrow , apoptosis \uparrow , differentiation \uparrow [3, 149, 156]
Linoleic acid	PPARs [20, 107, 108, 151]	Lipid synthesis \uparrow , differentiation \uparrow , conversion of testosterone to 5- α -dihydrotestosterone (5- α -DHT) \uparrow [20, 107, 108]
Testosterone, 5 α -DHT	Testosterone (androgen) receptor [22, 35, 95] (modification: 3 β -hydroxysteroid dehydrogenase/ Δ 5-4-isomerase, 17 β -hydroxysteroid dehydrogenase, 5 α -reductase-1 and 3 β -hydroxysteroid dehydrogenase) [73]	Proliferation \uparrow [35, 171] in the presence of cofactors (e.g., PPAR agonists): lipid synthesis \uparrow , differentiation \uparrow [107, 108]
Estrogens	Estrogen receptor- α and β [95]	Questionable (sebogenesis \downarrow ?) [26, 44]
Progesterone	Progesterone receptor [95]	?
(Glucocorticosteroids)	Glucocorticoid receptor (?)	Permissive effect [101], proliferation \uparrow , lipid synthesis \downarrow [18, 172]
Calcitriol (vitamin D ₃)	Vitamin D receptor (VDR) (modification: 1 α , 24- and 25-hydroxylases) [57]	IL-6 \downarrow , IL-8 \downarrow , cathelicidin \uparrow in rapidly proliferating cultures (with serum): proliferation \downarrow , cell cycle arrest in slowly proliferating cultures (without serum): proliferation \uparrow , lipid synthesis \downarrow [57, 63]
Growth hormone (GH)	Growth hormone receptor (GHR) [67, 85]	Differentiation \uparrow , no effect on proliferation [25]
Insulin-like growth factor-I (IGF-I)	IGF-I receptor (?) \rightarrow phosphoinositol-3-kinase/Akt (PI3K/Akt) \rightarrow sterol response element-binding protein-1 (SREBP1) [136, 137]	Proliferation \uparrow , minor effect on differentiation [25] lipid synthesis \uparrow [72]
Insulin	Insulin receptor	Proliferation \uparrow , differentiation \uparrow , supportive role in the effect of 5 α -DHT, GH, and IGF-I [25]
Epidermal growth factor (EGF)	EGF receptor (EGFR) [82]	Differentiation \downarrow , proliferation \uparrow [44]
Fibroblast growth factor-7 (FGF7)	Fibroblast growth factor receptor-2b (FGFR2b) [41, 78]	Acne formation \uparrow FGFR2b knockout mice show sebaceous gland atrophy [41, 78]
IL-1 β	Not described on sebocytes	IL-8 \uparrow [170]
β -endorphin	μ opioid receptor [12]	Lipid synthesis \uparrow , proliferation \downarrow , differentiation \uparrow [12]
Corticotropin-releasing hormone (CRH)	CRH receptor-1, 2 (CRHR1, CRHR2) [125, 170]	Lipid synthesis \uparrow , 3 β -hydroxysteroid dehydrogenase/ Δ 5-4 isomerase expression \uparrow , differentiation \uparrow , proliferation \downarrow , IL-6 \uparrow , IL-8 \uparrow [58, 170]
α -melanocyte-stimulating hormone (melanocortin, α MSH)	Melanocortin receptor-1, 5 (MC-1R, MC-5R) [13, 140, 145, 158, 159]	IL-8 \downarrow , differentiation \uparrow , lipid synthesis \uparrow [12, 13, 158, 159]
Adrenocorticotrophic hormone (corticotropin, ACTH)	Melanocortin receptor-2 (MC-2R) [43]	Differentiation \uparrow , lipid synthesis \uparrow [158]
Substance P (SP)	Not described on sebocytes	IL-1 \uparrow , IL-6 \uparrow , TNF- α \uparrow , PPAR γ \uparrow , lipid synthesis \uparrow , Sebaceous gland size \uparrow , differentiation \uparrow [64, 150]
Endocannabinoids (anandamide—AEA, 2-arachidonoyl glycerol—2-AG)	Cannabinoid receptor 2 (CB2) \rightarrow mitogen-activated protein kinase (MAPK) \rightarrow PPAR [31]	Lipid synthesis \uparrow , apoptosis \uparrow , differentiation \uparrow [31]
Capsaicin	Transient receptor potential vanilloid 1 (TRPV1) \rightarrow PPARs \downarrow and retinoid-X receptors (RXRs) \downarrow , extracellular Ca ²⁺ dependent [149]	Lipid synthesis \downarrow , differentiation \downarrow , IL-1 β \downarrow , proliferation \uparrow (TRPV1 specific), necrosis \uparrow (large dose, TRPV1 independent) [149]

The presence of epidermal growth factor (EGF) receptor was also identified on human sebocytes [82]. In animal models, EGF increased the number of cells in the sebaceous glands [74] whereas in human *in vitro* systems, it apparently inhibited sebaceous differentiation [44]. This latter finding was further supported by the observation that the most frequent side effects of the EGF receptor inhibitor monoclonal antibody cetuximab, used in cancer therapy, are the acneiform eruptions [148]. Besides EGF, recent results suggest the role of fibroblast growth factor receptor-2b (FGFR2b)-coupled signaling in the control of sebaceous functions and development of acne [41, 78].

Neuroendocrine regulators

In the last decade, a large number of endocrine mediators and neurotransmitters were reported to have significant influence on sebaceous gland biology (Table 1). Indeed, the skin expresses ligands and receptors for a broad range of neurohormones of the hypothalamic–pituitary axis [116, 127, 129, 163]. Various cell types of mouse and human skin express CRH receptors (CRHR1 and CRHR2) [105, 118, 119]. Likewise, these receptors were also detected in the human sebaceous gland [125]. Intriguingly, the skin is also able to produce the ligands of CRHRs: the expression of both CRH and urocortin was demonstrated in various cutaneous cell types, including those of the sebaceous gland [55, 120, 126]. Likewise, on cultured SZ95 sebocytes, both CRHR1 and CRHR2 as well as CRH were described [170] and the “autocrine” CRH, by acting on its receptors, was implicated to increase lipid synthesis and androgenic hormone production (by elevating the expression of 3β -hydroxysteroid dehydrogenase/ $\Delta 5$ -4 isomerase) [170]. Furthermore, CRH stimulated IL-6 and IL-8 release without affecting IL-1 α and IL-1 β release and, like urocortin, inhibited the proliferation of the SZ95 cells [58].

Of further importance, CRH also regulates the cleavage of proopiomelanocortin (POMC), whose presence was also reported in several cell types of mouse and human skin [118, 124], including sebocytes [56, 75]. Similar to the pituitary gland, the human skin is also able to process POMC to shorter POMC-derived peptides [155]. These peptides, such as α - and β -MSH (melanocortin), adrenocorticotrophic hormone (corticotropin, ACTH), and β -endorphin were demonstrated in sebocytes, and the presence of the prohormone convertase enzymes responsible for enzymatic cleavage of POMC was also documented [55, 75, 124]. Furthermore, the cutaneous expression of their receptors—melanocortin receptor types 1 and 2 (ACTH receptor) and type 5 (MC-1R, MC-2R, and MC-5R, respectively), and μ opioid receptor—were also described [13, 43, 140, 145, 158, 162].

Similar to CRH, POMC-derived peptides were also found to be active on sebocytes. MC-5R-coupled signaling

was shown to play a stimulatory role in secretion of mouse exocrine glands, including sebaceous glands [17]. POMC-derived peptides, such as MSH and ACTH, stimulated differentiation of human sebocytes which effect was correlated with an increased expression of MC-5R [158]. Melanocortin and its analogues were suggested to decrease IL-8 secretion and induce lipid synthesis via MC-1R on non-differentiated sebocytes, and via MC-5R on differentiated cells [12, 13, 159]. Pilot results also indicated that β -endorphin (most probably via acting on μ opioid receptors) exerts a differentiation-promoting effect; namely, it increased lipid synthesis and decreases proliferation of cultured sebocytes [12].

These intriguing data unambiguously argue for the presence of a functional hypothalamic–pituitary axis in the skin which may, therefore, play a key role in the regulation of cutaneous stress responses (reviewed in [128, 130]). However, the system may have distinct effects on different cutaneous cell types involving, e.g., both anti- and pro-inflammatory mechanisms [117]. For examples, in sebocytes, an increased expression of CRH, CRHRs, and CRH-binding protein was found in acne-prone skin compared to non-affected skin samples [38].

Biology of sebocytes may additionally be influenced by other paracrine mediators, neurotransmitters, and neuropeptides. Besides stimulating the release of pro-inflammatory cytokines (see above) [64], the sensory neuron-derived SP was found to accelerate differentiation and proliferation of sebaceous glands [150]. On SZ95 sebocytes, on which the presence of H1 histamine receptor was identified, anti-histamines were shown to decrease squalene synthesis [94]. Different muscarinic and nicotinic cholinergic receptor subunits were also found on non-differentiated and differentiated cells of sebaceous gland [60]. It is tempting to hypothesize, therefore, that the activation of these receptors by neural and paracrine acetylcholine or by nicotine from cigarette smoke may play a role in the pathogenesis of acne [165]. A few independent studies and observations furthermore suggest a potential influence of somatostatin, nerve growth factor, calcitonin gene-related peptide, neuropeptide-Y, and serotonin on sebocytes functions [12].

Hence, the plethora of these neuroendocrine agents and their receptors expressed on sebocytes may form a causative link between psycho-emotional stress and the development of acne [166].

Lipid mediators to control sebaceous lipid production

Besides the above wide array of regulatory mechanisms, the biology of sebaceous glands is strongly controlled by paracrine and autocrine lipid mediators. For example, AA

and linoleic acid were shown to induce terminal differentiation of sebocytes and induce lipid synthesis in cultured sebocytes [20, 156]. Therefore, besides the classical lipid-target nuclear receptors, in the sections below we introduce a few novel signaling pathways mediating or modifying the effects of these crucial lipid mediators (Table 1).

“Classical lipid targets”—role of nuclear receptors

PPARs, which belong to the thyroid-hormone receptor-like subfamily of nuclear receptors, are localized in the nucleus and form heterodimers with retinoid-X receptors (RXRs) [80]. PPARs play a central role in the regulation of lipid homeostasis of various tissues. Namely, PPARs can be activated by a wide array of lipid mediators, such as, e.g., fatty acids [27, 152], and PPAR γ became well-known as central regulator of adipocyte differentiation [40, 54, 61, 112].

Similar to adipocytes, sebocytes also exhibit an intensive lipid metabolism which is reportedly under the control of PPARs. Sebocytes express all three PPAR isoforms, PPAR α , PPAR γ , and PPAR δ ; among them, PPAR γ seems to be the dominant form whereas the expression of PPAR δ is low. Of great importance, the natural PPAR ligand linoleic acid (similar to synthetic agonists) was shown to increase the lipid production and differentiation of sebocytes [20, 108, 151]. The differentiation-promoting AA can also stimulate PPARs; likewise, the AA-derivative LTB₄ was shown to act as a potent activator of PPAR α [28]. Furthermore, PPARs take part in the regulation of other cellular processes of sebocytes. Namely, PPAR γ was found to be involved in the control of PGE₂ production [160] whereas the effects of androgens to modulate sebaceous functions are also mediated by PPARs [107, 108]. Recently, the vitamin D₃ was reported to increase the PPAR α expression of SZ95 cell line [115].

Recently, the presence of liver-X receptor (LXR) was also identified on human sebocytes [109]. Similar to PPARs, the LXR has a critical role in cholesterol homeostasis and lipid metabolism. Activation of LXR α on SZ95 sebocytes decreased proliferation, increased lipid synthesis, and induced the expression of LXR target genes, such as fatty acid synthase and sterol regulatory-binding protein-1. Furthermore, stimulation of LXR α down-regulated the expression of COX-2 and inducible nitric oxide synthase, induced by LPS treatment. Finally, similar to as reported on adipocytes, LXR activation increased the expression of PPARs on SZ95 sebocytes as well [47, 109].

Role of the endocannabinoid system and related TRP channels

Research efforts of the last two decades have unambiguously confirmed that the human body is able to produce

various molecules which exhibit similar biological effects to those agents which can be found in the “infamous” plant *Cannabis sativa*. These substances are the endogenous cannabinoids, among which the best known ones are the *N*-arachidonoyl ethanolamine (anandamide) and 2-arachidonoyl glycerol (2-AG) [71, 77]. Endocannabinoids may target various receptor structures, which include the G-protein-coupled “classical” CB1 and CB2 cannabinoid receptors; the “novel”, also G-protein-coupled GPR55, GPR119, and GPR18; as well as the “ionotropic” cannabinoid receptors, which are certain members of the transient receptor potential (TRP) ion channel superfamily, namely TRPV1, TRPV2, TRPV4, TRPA1, and TRPM8 [2, 14, 24, 49, 50, 71, 76]. Furthermore, certain endocannabinoids may also influence additional signal transduction systems by modulating the activity of other receptors, e.g., ionotropic glutamate receptors, nicotinic acetylcholine receptors, serotonin receptors, μ opioid receptors, and different PPARs [15, 24, 91, 96, 98]. Of further importance, endocannabinoid synthesizing (*N*-acyl-phosphatidylethanolamine-specific phospholipase D and diacylglycerol lipases [DAGL α and β]) and degrading enzymes (fatty acid amide hydrolase and monoacylglycerol lipase) as well as anandamide/endocannabinoid membrane transporters were also described [23, 29, 34]. Therefore, the endocannabinoids, their receptors, and the enzymes involved in the endocannabinoid metabolism are collectively referred to as the endocannabinoid system (ECS), one of the most complex signaling systems of the human body. Indeed, ECS regulates various biological processes of the human body such as, e.g., food intake, energy balance, body mass, memory, immunological, and vascular responses, bone metabolism, endocrine homeostasis, etc. [70, 71, 92].

Intriguingly, functional ECS has lately been identified in the skin as well, and was implicated in key regulatory processes affecting cutaneous biology [7]. Several human skin cell compartments such as, e.g., epidermal and hair follicle keratinocytes and sebaceous gland-derived sebocytes were shown to produce prototypic endocannabinoids, and express metabotropic and ionotropic cannabinoid receptors and metabolic enzymes [9–11, 16, 31, 51, 53, 69, 142, 143, 149]. Furthermore, stimulation of CB1 by anandamide (which can be produced by the hair follicles themselves) inhibited in vitro hair shaft elongation and induced apoptosis-driven premature catagen regression of the hair follicle [143].

With respect to sebaceous gland biology, it is also of great importance that anandamide and 2-AG are produced by human sebaceous gland-derived SZ95 sebocytes which predominantly express CB2 (Table 1). Both endocannabinoids stimulated lipid production via CB2-coupled signaling involving the MAPK pathway and the up-regulation of PPARs. Since cells with “silenced” CB2 exhibited signif-

icantly suppressed basal lipid production, these results collectively suggest that human sebocytes utilize an autocrine/paracrine, endogenously (and most probably constitutively) active, CB2-mediated ECS for positively regulating lipid production [31] (Fig. 1).

Sebocytes also express “ionotropic” cannabinoid receptors. Among these, the “capsaicin-receptor” TRPV1—which was shown to be activated by anandamide on various cells types including, e.g., sensory neurons [30, 173]—was identified both in situ on human sebaceous glands [11, 141] and in vitro on cultured SZ95 sebocytes [149]. However, in contrast to previous findings, anandamide does not seem to activate TRPV1 on sebocytes. Actually, TRPV1-coupled signaling evoked opposite effects

to those induced by anandamide since the stimulation of TRPV1 by capsaicin inhibited both basal and AA-induced lipid synthesis in an extracellular calcium-dependent manner [149]. Moreover, in parallel to the inhibition of sebum production, capsaicin treatment down-regulated the expressions of PPAR and related RXR isoforms in SZ95 sebocytes [149]. These results collectively argue for that TRPV1 signaling (in contrast to the action of the sebaceous ECS) inhibits terminal differentiation of sebocytes.

Intriguingly, preliminary findings suggest the possible involvement of other TRP channels in sebaceous gland biology. Besides TRPV1, the presence of TRPV2, TRPV3, and TRPV4 was also identified on SZ95 sebocytes [89, 90]. Moreover, certain plant derived and synthetic

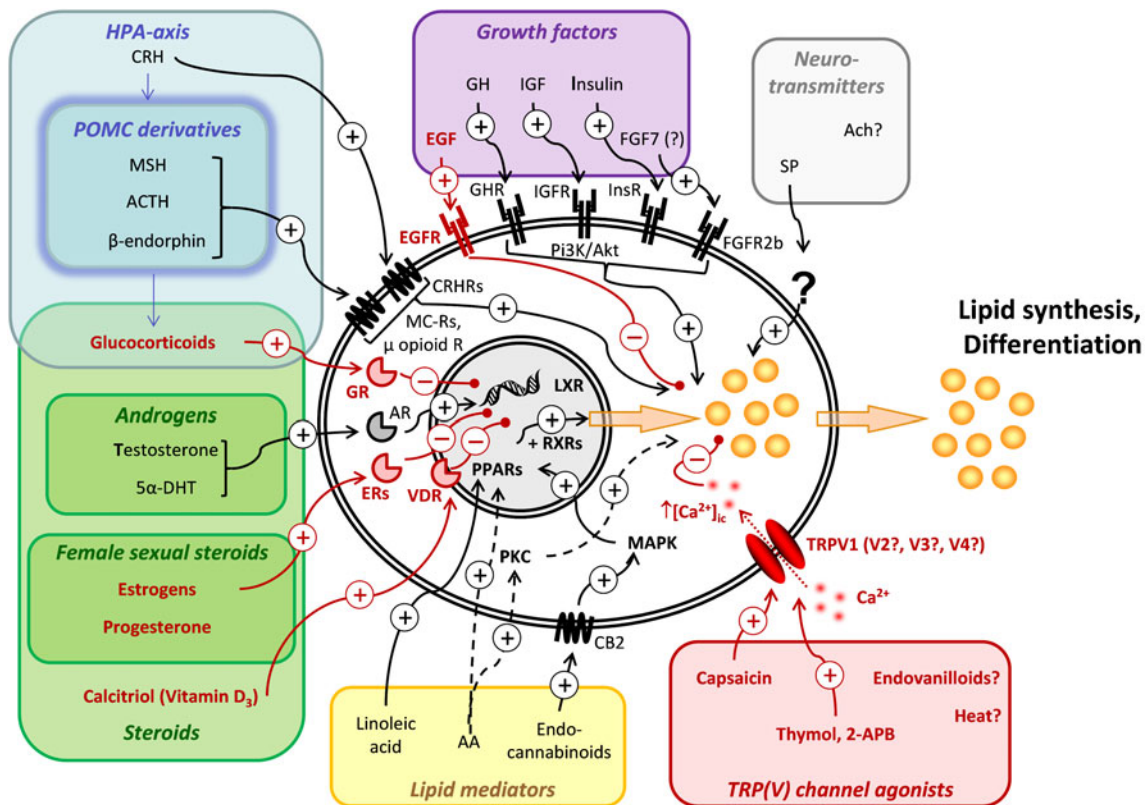


Fig. 1 Control of sebocyte differentiation. Sebaceous lipid synthesis can be stimulated (black letters and arrows) and inhibited (red letters and arrows) by various mediators/agents and related signaling pathways. These agents activate several surface membrane receptors involving both G-protein-coupled ones and receptor tyrosine kinases, as well as certain nuclear receptors. In the differentiation processes, expression and/or activation of certain “lipid genes” (e.g., PPAR transcription factors) play pivotal roles. These genes can either be activated directly (e.g., by certain lipid mediators) or indirectly via intracellular signaling pathways (e.g., MAPK system). Intriguingly, elevation of intracellular calcium concentration in sebocytes by the stimulation of TRPV1 (and most probably other TRPV channels) appears to inhibit the lipogenic actions of the above mediators. These novel mechanisms may be exploited in the clinical management of sebaceous gland diseases with altered sebum production (e.g., acne vulgaris and dry skin conditions). *Abbreviations:* HPA hypothalamic–

pituitary–adrenal cortex, CRH corticotropin-releasing hormone, POMC proopiomelanocortin, MSH melanocyte stimulating hormone, melanocortin, ACTH adrenocorticotrophic hormone, corticotropin, 5 α -DHT 5 α -dihydrotestosterone, EGF epidermal growth factor, EGFR EGF receptor, GH growth hormone, GHR GH receptor, IGF insulin-like growth factor, IGFR IGF receptor, InsR insulin receptor, FGF7 fibroblast growth factor-7, FGFR2b FGF receptor-2b, SP substance P, Ach acetylcholine, Pi3K phosphoinositol-3-kinase, CRHRs CRH receptors, MC-Rs melanocortin receptors, GR glucocorticoid receptor, AR androgen receptor, ERs estrogen receptors, VDR vitamin D receptor, PPARs peroxisome proliferator-activated receptors, LXR liver-X receptor, RXRs retinoid-X receptors, PKC protein kinase C, MAPK mitogen-activated protein kinase, $[Ca^{2+}]_{ic}$ intracellular Ca^{2+} concentration, TRPV1 transient receptor potential vanilloid 1, CB2 cannabinoid receptor subtype 2, AA arachidonic acid, 2-APB 2-aminoethoxydiphenyl borate

activators of these channels (e.g., thymol, eugenol, 2-aminoethoxydiphenyl borate) were able to induce transient elevation of the intracellular calcium concentration ($[Ca^{2+}]_{ic}$) and suppressed sebum production induced by AA treatment (similar to the action of TRPV1 stimulation). At present, it is unclear how specific these effects are as dramatic increases in calcium concentration will likely affect proliferation and differentiation of sebocytes, similar to other human skin cells [79, 157]. Indeed, previous reports suggest that the differentiation of sebocytes could be stimulated by the decrease of the extracellular Ca^{2+} concentration [114]. Taken together, it can be postulated that the elevation of $[Ca^{2+}]_{ic}$ by TRPV activation may inhibit differentiation and the closely related lipid production in human sebaceous gland cells (Fig. 1).

Concluding remarks—“Bite the dog that bit you”: lipids that target sebaceous diseases

The above, most recently discovered regulatory mechanisms introduce novel therapeutic strategies to manage certain sebaceous gland disorders. A group of these disorders can be characterized by hyperactivity of the sebaceous glands, and associated with overproduction of sebum and inflammatory processes. In other diseases, conversely, the hypofunction of the gland may lead to the development of such conditions as dry skin and associated syndromes. In some rare cases, the hyperproliferation of sebaceous gland may result in sebaceous tumor formation.

Among these disorders, doubtless, acne vulgaris has the highest prevalence. Classical strategies in treatment of acne involves antibiotic treatment, oral contraceptives, and isotretinoin (13(cis)-retinoic acid). Targeting the presented novel regulatory mechanisms may broaden the therapeutic arsenal with anti-androgens, 5- α -reductase inhibitors, LOX, and/or COX inhibitors, and insulin-sensitizing agents acting on PPARs [59].

In addition, recent findings focus the attention on the influence of novel lipid-signaling mechanisms. Among these, the targeted manipulation of the sebaceous ECS may be an effective strategy. For example, in acne and other inflammatory sebaceous gland diseases characterized by sebaceous hyperfunction, the inhibition of CB2-mediated signaling may be of therapeutic value. Alternatively, inhibition of endocannabinoid synthesizing or stimulation of degrading enzymes (thereby “suppressing the sebaceous endocannabinoid tone”) may also be beneficial. Conversely, in diseases associated with dry skin conditions or sebaceous hyperproliferation, the activation of endocannabinoid signaling or “augmenting the ECS tone” may be preferable [7]. Furthermore, activators of certain TRPV channels may also have the desired sebostatic

effect in acne [8]. Since transdermal penetration of most of these molecules is well established, it can be envisaged these agents could be efficiently applied topically to the skin in the form of a cream.

Taken together, future clinical studies are now warranted to explore the real therapeutic potential of these intriguing (mostly pre-clinical) data.

Acknowledgment This work was supported in part by Hungarian (OTKA NK78398, OTKA NNF78456, TÁMOP-4.2.2-08/1/2008-0019, TÁMOP 4.2.1/B-09/1/KONV-2010-0007) and EU (FP7-REGPOT-2008-1/22992) research grants. GC is a recipient of the János Bolyai scholarship of the Hungarian Academy of Sciences. OA is recipient of the “Richter Talentum” scholarship.

References

1. Akamatsu H, Zouboulis CC, Orfanos CE (1992) Control of human sebocyte proliferation in vitro by testosterone and 5-alpha-dihydrotestosterone is dependent on the localization of the sebaceous glands. *J Invest Dermatol* 99:509–511
2. Akopian AN, Ruparel NB, Jeske NA, Patwardhan A, Hargreaves KM (2009) Role of ionotropic cannabinoid receptors in peripheral antinociception and antihyperalgesia. *Trends Pharmacol Sci* 30:79–84
3. Alestas T, Ganceviciene R, Fimmel S, Muller-Decker K, Zouboulis CC (2006) Enzymes involved in the biosynthesis of leukotriene B4 and prostaglandin E2 are active in sebaceous glands. *J Mol Med* 84:75–87
4. Allen M, Grachtchouk M, Sheng H, Grachtchouk V, Wang A, Wei LB, Liu JH, Ramirez A, Metzger D, Chambon P, Jorcano J, Dlugosz AA (2003) Hedgehog signaling regulates sebaceous gland development. *Am J Pathol* 163:2173–2178
5. Bikle DD (1995) 1, 25(OH)2D3-regulated human keratinocyte proliferation and differentiation: basic studies and their clinical application. *J Nutr* 125:1709S–1714S
6. Bikle DD, Oda Y, Xie Z (2004) Calcium and 1, 25(OH)2D: interacting drivers of epidermal differentiation. *J Steroid Biochem Mol Biol* 89–90:355–360
7. Biro T, Toth BI, Hasko G, Paus R, Pacher P (2009) The endocannabinoid system of the skin in health and disease: novel perspectives and therapeutic opportunities. *Trends Pharmacol Sci* 30:411–420
8. Biro T, Toth BI, Marincsak R, Dobrosi N, Geczy T, Paus R (2007) TRP channels as novel players in the pathogenesis and therapy of itch. *Biochim Biophys Acta* 1772:1004–1021
9. Blazquez C, Carracedo A, Barrado L, Real PJ, Fernandez-Luna JL, Velasco G, Malumbres M, Guzman M (2006) Cannabinoid receptors as novel targets for the treatment of melanoma. *FASEB J* 20:2633–2635
10. Bodo E, Biro T, Telek A, Czifra G, Griger Z, Toth BI, Mescalchin A, Ito T, Bettermann A, Kovacs L, Paus R (2005) A hot new twist to hair biology: involvement of vanilloid receptor-1 (VR1/TRPV1) signaling in human hair growth control. *Am J Pathol* 166:985–998
11. Bodo E, Kovacs I, Telek A, Varga A, Paus R, Kovacs L, Biro T (2004) Vanilloid receptor-1 (VR1) is widely expressed on various epithelial and mesenchymal cell types of human skin. *J Invest Dermatol* 123:410–413
12. Bohm M (2009) Neuroendocrine regulators: novel trends in sebaceous gland research with future perspectives for the

- treatment of acne and related disorders. *Dermatoendocrinol* 1:136–140
13. Bohm M, Schiller M, Stander S, Seltmann H, Li Z, Brzoska T, Metz D, Schioth HB, Skottner A, Seiffert K, Zouboulis CC, Luger TA (2002) Evidence for expression of melanocortin-1 receptor in human sebocytes in vitro and in situ. *J Invest Dermatol* 118:533–539
 14. Brown AJ (2007) Novel cannabinoid receptors. *Br J Pharmacol* 152:567–575
 15. Burstein S (2005) PPAR-gamma: a nuclear receptor with affinity for cannabinoids. *Life Sci* 77:1674–1684
 16. Casanova ML, Blazquez C, Martinez-Palacio J, Villanueva C, Fernandez-Acenero MJ, Huffman JW, Jorcano JL, Guzman M (2003) Inhibition of skin tumor growth and angiogenesis in vivo by activation of cannabinoid receptors. *J Clin Invest* 111:43–50
 17. Chen W, Kelly MA, Opitz-Araya X, Thomas RE, Low MJ, Cone RD (1997) Exocrine gland dysfunction in MC5-R-deficient mice: evidence for coordinated regulation of exocrine gland function by melanocortin peptides. *Cell* 91:789–798
 18. Chen W, Liao CY, Hung CL, Lin TK, Sheu HM, Zouboulis CC (2006) Potent corticosteroids inhibit lipogenesis in sebaceous glands. *Dermatology* 213:264–265
 19. Chen W, Thiboutot D, Zouboulis CC (2002) Cutaneous androgen metabolism: basic research and clinical perspectives. *J Invest Dermatol* 119:992–1007
 20. Chen W, Yang CC, Sheu HM, Seltmann H, Zouboulis CC (2003) Expression of peroxisome proliferator-activated receptor and CCAAT/enhancer binding protein transcription factors in cultured human sebocytes. *J Invest Dermatol* 121:441–447
 21. Chen WC, Zouboulis CC (2009) Hormones and the pilosebaceous unit. *Dermatoendocrinol* 1:81–86
 22. Choudhry R, Hodgins MB, Van der Kwast TH, Brinkmann AO, Boersma WJ (1992) Localization of androgen receptors in human skin by immunohistochemistry: implications for the hormonal regulation of hair growth, sebaceous glands and sweat glands. *J Endocrinol* 133:467–475
 23. De Petrocellis L, Cascio MG, Di Marzo V (2004) The endocannabinoid system: a general view and latest additions. *Br J Pharmacol* 141:765–774
 24. Demuth DG, Molleman A (2006) Cannabinoid signalling. *Life Sci* 78:549–563
 25. Deplewski D, Rosenfield RL (1999) Growth hormone and insulin-like growth factors have different effects on sebaceous cell growth and differentiation. *Endocrinology* 140:4089–4094
 26. Deplewski D, Rosenfield RL (2000) Role of hormones in pilosebaceous unit development. *Endocr Rev* 21:363–392
 27. Desvergne B, Wahli W (1999) Peroxisome proliferator-activated receptors: nuclear control of metabolism. *Endocr Rev* 20:649–688
 28. Devchand PR, Keller H, Peters JM, Vazquez M, Gonzalez FJ, Wahli W (1996) The PPARalpha-leukotriene B4 pathway to inflammation control. *Nature* 384:39–43
 29. Di Marzo V (2008) Endocannabinoids: synthesis and degradation. *Rev Physiol Biochem Pharmacol* 160:1–24
 30. Dinis P, Charrua A, Avelino A, Yaqoob M, Bevan S, Nagy I, Cruz F (2004) Anandamide-evoked activation of vanilloid receptor 1 contributes to the development of bladder hyperreflexia and nociceptive transmission to spinal dorsal horn neurons in cystitis. *J Neurosci* 24:11253–11263
 31. Dobrosi N, Toth BI, Nagy G, Dozsa A, Geczy T, Nagy L, Zouboulis CC, Paus R, Kovacs L, Biro T (2008) Endocannabinoids enhance lipid synthesis and apoptosis of human sebocytes via cannabinoid receptor-2-mediated signaling. *FASEB J* 22:3685–3695
 32. Drake DR, Brogden KA, Dawson DV, Wertz PW (2008) Thematic review series: skin lipids. Antimicrobial lipids at the skin surface. *J Lipid Res* 49:4–11
 33. Farrar MD, Ingham E (2004) Acne: inflammation. *Clin Dermatol* 22:380–384
 34. Fowler CJ, Jacobsson SO (2002) Cellular transport of anandamide, 2-arachidonoylglycerol and palmitoylethanolamide—targets for drug development? *Prostaglandins Leukot Essent Fatty Acids* 66:193–200
 35. Fritsch M, Orfanos CE, Zouboulis CC (2001) Sebocytes are the key regulators of androgen homeostasis in human skin. *J Invest Dermatol* 116:793–800
 36. Fujie T, Shikiji T, Uchida N, Urano Y, Nagae H, Arase S (1996) Culture of cells derived from the human sebaceous gland under serum-free conditions without a biological feeder layer or specific matrices. *Arch Dermatol Res* 288:703–708
 37. Funk CD (2001) Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science* 294:1871–1875
 38. Ganceviciene R, Graziene V, Fimmel S, Zouboulis CC (2009) Involvement of the corticotropin-releasing hormone system in the pathogenesis of acne vulgaris. *Br J Dermatol* 160:345–352
 39. Georgel P, Crozat K, Lauth X, Makrantonaki E, Seltmann H, Sovath S, Hoebe K, Du X, Rutschmann S, Jiang Z, Bigby T, Nizet V, Zouboulis CC, Beutler B (2005) A toll-like receptor 2-responsive lipid effector pathway protects mammals against skin infections with gram-positive bacteria. *Infect Immun* 73:4512–4521
 40. Gregoire FM, Smas CM, Sul HS (1998) Understanding adipocyte differentiation. *Physiol Rev* 78:783–809
 41. Grose R, Fantl V, Werner S, Chioni AM, Jarosz M, Rudling R, Cross B, Hart IR, Dickson C (2007) The role of fibroblast growth factor receptor 2b in skin homeostasis and cancer development. *EMBO J* 26:1268–1278
 42. Guha U, Mecklenburg L, Cowin P, Kan L, O'Guin WM, D'Vizio D, Pestell RG, Paus R, Kessler JA (2004) Bone morphogenetic protein signaling regulates postnatal hair follicle differentiation and cycling. *Am J Pathol* 165:729–740
 43. Guo HW, Deng J, Yang XC, Zhong BY, Shen Z, Yang SY, Liu BH, Hao F (2010) Melanocortin receptor type 2 (MC2R, ACTH receptor) expression in patients with alopecia areata. *Exp Dermatol* 19:1020–1022
 44. Guy R, Ridden C, Kealey T (1996) The improved organ maintenance of the human sebaceous gland: modeling in vitro the effects of epidermal growth factor, androgens, estrogens, 13-cis retinoic acid, and phenol red. *J Invest Dermatol* 106:454–460
 45. Han GW, Li AG, Liang YY, Owens P, He W, Lu SL, Yoshimatsu Y, Wang D, ten Dijke P, Lin X, Wang XJ (2006) Smad7-induced beta-catenin degradation alters epidermal appendage development. *Dev Cell* 11:301–312
 46. Holick MF, Chen ML, Kong XF, Sanan DK (1996) Clinical uses for calcitropic hormones 1, 25-dihydroxyvitamin D3 and parathyroid hormone-related peptide in dermatology: a new perspective. *J Investig Dermatol Symp Proc* 1:1–9
 47. Hong I, Lee MH, Na TY, Zouboulis CC, Lee MO (2008) LXR alpha enhances lipid synthesis in SZ95 sebocytes. *J Investig Dermatol* 128:1266–1272
 48. Horsley V, O'Carroll D, Tooze R, Ohinata Y, Saitou M, Obukhanych T, Nussenzweig M, Tarakhovskiy A, Fuchs E (2006) Blimp1 defines a progenitor population that governs cellular input to the sebaceous gland. *Cell* 126:597–609
 49. Howlett AC (2005) Cannabinoid receptor signaling. *Handb Exp Pharmacol* 168:53–79
 50. Howlett AC, Barth F, Bonner TI, Cabral G, Casellas P, Devane WA, Felder CC, Herkenham M, Mackie K, Martin BR, Mechoulam R, Pertwee RG (2002) International Union of Pharmacology. XXVII. Classification of cannabinoid receptors. *Pharmacol Rev* 54:161–202

51. Ibrahim MM, Porreca F, Lai J, Albrecht PJ, Rice FL, Khodorova A, Davar G, Makriyannis A, Vanderah TW, Mata HP, Malan TP Jr (2005) CB2 cannabinoid receptor activation produces antinociception by stimulating peripheral release of endogenous opioids. *Proc Natl Acad Sci U S A* 102:3093–3098
52. Ito N, Ito T, Kromminga A, Bettermann A, Takigawa M, Kees F, Straub RH, Paus R (2005) Human hair follicles display a functional equivalent of the hypothalamic–pituitary–adrenal axis and synthesize cortisol. *FASEB J* 19:1332–1334
53. Karsak M, Gaffal E, Date R, Wang-Eckhardt L, Rehnelt J, Petrosino S, Starowicz K, Steuder R, Schlicker E, Cravatt B, Mechoulam R, Buettner R, Werner S, Di Marzo V, Tuting T, Zimmer A (2007) Attenuation of allergic contact dermatitis through the endocannabinoid system. *Science* 316:1494–1497
54. Kersten S, Desvergne B, Wahli W (2000) Roles of PPARs in health and disease. *Nature* 405:421–424
55. Kim HS, Cho DH, Kim HJ, Lee JY, Cho BK, Park HJ (2006) Immunoreactivity of corticotropin-releasing hormone, adrenocorticotrophic hormone and alpha-melanocyte-stimulating hormone in alopecia areata. *Exp Dermatol* 15:515–522
56. Kono M, Nagata H, Umemura S, Kawana S, Osamura RY (2001) In situ expression of corticotropin-releasing hormone (CRH) and proopiomelanocortin (POMC) genes in human skin. *FASEB J* 15:2297–2299
57. Kramer C, Seltmann H, Seifert M, Tilgen W, Zouboulis CC, Reichrath J (2009) Characterization of the vitamin D endocrine system in human sebocytes in vitro. *J Steroid Biochem Mol Biol* 113:9–16
58. Krause K, Schnitger A, Fimmel S, Glass E, Zouboulis CC (2007) Corticotropin-releasing hormone skin signaling is receptor-mediated and is predominant in the sebaceous glands. *Horm Metab Res* 39:166–170
59. Kurokawa I, Danby FW, Ju Q, Wang X, Xiang LF, Xia L, Chen W, Nagy I, Picardo M, Suh DH, Ganceviciene R, Schagen S, Tsatsou F, Zouboulis CC (2009) New developments in our understanding of acne pathogenesis and treatment. *Exp Dermatol* 18:821–832
60. Kurzen H, Berger H, Jager C, Hartschuh W, Naher H, Gratchev A, Goerdts S, Deichmann M (2004) Phenotypical and molecular profiling of the extraneuronal cholinergic system of the skin. *J Invest Dermatol* 123:937–949
61. Lee CH, Olson P, Evans RM (2003) Minireview: lipid metabolism, metabolic diseases, and peroxisome proliferator-activated receptors. *Endocrinology* 144:2201–2207
62. Lee DY, Huang CM, Nakatsuji T, Thiboutot D, Kang SA, Monestier M, Gallo RL (2009) Histone H4 is a major component of the antimicrobial action of human sebocytes. *J Invest Dermatol* 129:2489–2496
63. Lee DY, Yamasaki K, Rudsil J, Zouboulis CC, Park GT, Yang JM, Gallo RL (2008) Sebocytes express functional cathelicidin antimicrobial peptides and can act to kill propionibacterium acnes. *J Invest Dermatol* 128:1863–1866
64. Lee WJ, Jung HD, Lee HJ, Kim BS, Lee SJ, do Kim W (2008) Influence of substance-P on cultured sebocytes. *Arch Dermatol Res* 300:311–316
65. Leveque JL, Pierard-Franchimont C, de Rigal J, Saint-Leger D, Pierard GE (1991) Effect of topical corticosteroids on human sebum production assessed by two different methods. *Arch Dermatol Res* 283:372–376
66. Lo Celso C, Berta MA, Braun KM, Frye M, Lyle S, Zouboulis CC, Watt FM (2008) Characterization of bipotential epidermal progenitors derived from human sebaceous gland: contrasting roles of c-Myc and beta-catenin. *Stem Cells* 26:1241–1252
67. Lobie PE, Breipohl W, Lincoln DT, Garcia-Aragon J, Waters MJ (1990) Localization of the growth hormone receptor/binding protein in skin. *J Endocrinol* 126:467–471
68. Lupi O (2008) Ancient adaptations of human skin: why do we retain sebaceous and apocrine glands? *Int J Dermatol* 47:651–654
69. Maccarrone M, Di Rienzo M, Battista N, Gasperi V, Guerrieri P, Rossi A, Finazzi-Agro A (2003) The endocannabinoid system in human keratinocytes. Evidence that anandamide inhibits epidermal differentiation through CB1 receptor-dependent inhibition of protein kinase C, activation protein-1, and transglutaminase. *J Biol Chem* 278:33896–33903
70. Mackie K (2006) Cannabinoid receptors as therapeutic targets. *Annu Rev Pharmacol Toxicol* 46:101–122
71. Mackie K, Brewer HB, Cota D, Cravatt BF, Di Marzo V, Ginsberg HN, Howlett A, Reggio PH, Woods SC (2008) The endocannabinoid system handbook. *Scientific*. Available via Endocannabinoid System Network (ECSN). <http://www.endocannabinoid.net/ECSHandbook/posttest/>
72. Makrantonaki E, Vogel K, Fimmel S, Oeff M, Seltmann H, Zouboulis CC (2008) Interplay of IGF-I and 17 beta-estradiol at age-specific levels in human sebocytes and fibroblasts in vitro. *Exp Gerontol* 43:939–946
73. Makrantonaki E, Zouboulis CC (2007) Testosterone metabolism to 5alpha-dihydrotestosterone and synthesis of sebaceous lipids is regulated by the peroxisome proliferator-activated receptor ligand linoleic acid in human sebocytes. *Br J Dermatol* 156:428–432
74. Matias JR, Orentreich N (1983) Stimulation of hamster sebaceous glands by epidermal growth factor. *J Invest Dermatol* 80:516–519
75. Mazurkiewicz JE, Corliss D, Slominski A (2000) Spatiotemporal expression, distribution, and processing of POMC and POMC-derived peptides in murine skin. *J Histochem Cytochem* 48:905–914
76. McHugh D, Hu SS, Rimmerman N, Juknat A, Vogel Z, Walker JM, Bradshaw HB (2010) N-arachidonoyl glycine, an abundant endogenous lipid, potently drives directed cellular migration through GPR18, the putative abnormal cannabidiol receptor. *BMC Neurosci* 11:44
77. Mechoulam R, Frideri E, Di Marzo V (1998) Endocannabinoids. *Eur J Pharmacol* 359:1–18
78. Melnik BC, Schmitz G, Zouboulis CC (2009) Anti-acne agents attenuate FGFR2 signal transduction in acne. *J Invest Dermatol* 129:1868–1877
79. Micallef L, Belaubre F, Pinon A, Jayat-Vignoles C, Delage C, Charveron M, Simon A (2009) Effects of extracellular calcium on the growth-differentiation switch in immortalized keratinocyte HaCaT cells compared with normal human keratinocytes. *Exp Dermatol* 18:143–151
80. Michalik L, Auwerx J, Berger JP, Chatterjee VK, Glass CK, Gonzalez FJ, Grimaldi PA, Kadowaki T, Lazar MA, O'Rahilly S, Palmer CN, Plutzky J, Reddy JK, Spiegelman BM, Staels B, Wahli W (2006) International Union of Pharmacology. LXI. Peroxisome proliferator-activated receptors. *Pharmacol Rev* 58:726–741
81. Nagy I, Pivarcsi A, Kis K, Koreck A, Bodai L, McDowell A, Seltmann H, Patrick S, Zouboulis CC, Kemeny L (2006) Propionibacterium acnes and lipopolysaccharide induce the expression of antimicrobial peptides and proinflammatory cytokines/chemokines in human sebocytes. *Microbes Infect* 8:2195–2205
82. Nanney LB, Stoscheck CM, King LE Jr, Underwood RA, Holbrook KA (1990) Immunolocalization of epidermal growth factor receptors in normal developing human skin. *J Invest Dermatol* 94:742–748
83. Niemann C, Uden AB, Lyle S, Zouboulis CC, Toftgard R, Watt FM (2003) Indian hedgehog and beta-catenin signaling: role in

- the sebaceous lineage of normal and neoplastic mammalian epidermis. *Proc Natl Acad Sci U S A* 100:11873–11880
84. Nowak JA, Polak L, Pasolli HA, Fuchs E (2008) Hair follicle stem cells are specified and function in early skin morphogenesis. *Cell Stem Cell* 3:33–43
 85. Oakes SR, Haynes KM, Waters MJ, Herington AC, Werther GA (1992) Demonstration and localization of growth hormone receptor in human skin and skin fibroblasts. *J Clin Endocrinol Metab* 75:1368–1373
 86. Oeff MK, Seltmann H, Hakiy N, Bogdanoff B, Nastos A, Walters R, Bornstein SR, Zouboulis CC (2002) Toll-like receptor 2 and 4-dependent regulation of inflammatory signaling in human sebocytes. *J Invest Dermatol* 119:736–736
 87. Oeff MK, Seltmann H, Hiroi N, Nastos A, Makrantonaki E, Bornstein SR, Zouboulis CC (2006) Differential regulation of Toll-like receptor and CD14 pathways by retinoids and corticosteroids in human sebocytes. *Dermatology* 213:266
 88. Ohsawa K, Watanabe T, Matsukawa R, Yoshimura Y, Imaeda K (1984) The possible role of squalene and its peroxide of the sebum in the occurrence of sunburn and protection from the damage caused by U.V. irradiation. *J Toxicol Sci* 9:151–159
 89. Olah A, Toth BI, Czifra G, Zouboulis CC, Paus R, Kovacs L, Biro T (2009) Cannabidiol inhibits lipid synthesis in human sebaceous gland-derived sebocytes—is cannabidiol a novel anti-acne agent? *J Invest Dermatol* 129:S58–S58
 90. Olah A, Toth IB, Szollosi AG, Czifra G, Sugawara K, Zouboulis CC, Paus R, Biro T (2010) Endo- and phytocannabinoids differentially regulate biology of human epithelial skin cells. *J Invest Dermatol* 130:S107–S107
 91. O'Sullivan SE (2007) Cannabinoids go nuclear: evidence for activation of peroxisome proliferator-activated receptors. *Br J Pharmacol* 152:576–582
 92. Pacher P, Batkai S, Kunos G (2006) The endocannabinoid system as an emerging target of pharmacotherapy. *Pharmacol Rev* 58:389–462
 93. Pappas A (2009) Epidermal surface lipids. *Dermatoendocrinol* 1:72–76
 94. Pelle E, McCarthy J, Seltmann H, Huang X, Mammone T, Zouboulis CC, Maes D (2008) Identification of histamine receptors and reduction of squalene levels by an antihistamine in sebocytes. *J Invest Dermatol* 128:1280–1285
 95. Pelletier G, Ren L (2004) Localization of sex steroid receptors in human skin. *Histol Histopathol* 19:629–636
 96. Pertwee RG (2005) Pharmacological actions of cannabinoids. *Handb Exp Pharmacol* 168:1–51
 97. Picardo M, Ottaviani M, Camera E, Mastrofrancesco A (2009) Sebaceous gland lipids. *Dermatoendocrinol* 1:68–71
 98. Pistis M, Melis M (2010) From surface to nuclear receptors: the endocannabinoid family extends its assets. *Curr Med Chem* 17:1450–1467
 99. Plewig G, Luderschmidt C (1977) Hamster ear model for sebaceous glands. *J Invest Dermatol* 68:171–176
 100. Pochi PE, Strauss JS (1977) Studies on the sebaceous glands in acne and endocrine disorders. *Bull N Y Acad Med* 53:359–367
 101. Pochi PE, Strauss JS, Mescon H (1963) The role of adrenocortical steroids in the control of human sebaceous gland activity. *J Invest Dermatol* 41:391–399
 102. Porter AM (2001) Why do we have apocrine and sebaceous glands? *J R Soc Med* 94:236–237
 103. Potter JE, Prutkin L, Wheatley VR (1979) Sebaceous gland differentiation. I. Separation, morphology and lipogenesis of isolated cells from the mouse preputial gland tumor. *J Invest Dermatol* 72:120–127
 104. Ro BI, Dawson TL (2005) The role of sebaceous gland activity and scalp microfloral metabolism in the etiology of seborrheic dermatitis and dandruff. *J Invest Dermatol Symp Proc* 10:194–197
 105. Roloff B, Fechner K, Slominski A, Furkert J, Botchkarev VA, Bulfone-Paus S, Zipper J, Krause E, Paus R (1998) Hair cycle-dependent expression of corticotropin-releasing factor (CRF) and CRF receptors in murine skin. *FASEB J* 12:287–297
 106. Rosenfield RL (1989) Relationship of sebaceous cell stage to growth in culture. *J Invest Dermatol* 92:751–754
 107. Rosenfield RL, Deplewski D, Kentsis A, Ciletti N (1998) Mechanisms of androgen induction of sebocyte differentiation. *Dermatology* 196:43–46
 108. Rosenfield RL, Kentsis A, Deplewski D, Ciletti N (1999) Rat preputial sebocyte differentiation involves peroxisome proliferator-activated receptors. *J Invest Dermatol* 112:226–232
 109. Russell LE, Harrison WJ, Bahta AW, Zouboulis CC, Burren JM, Philpott MP (2007) Characterization of liver X receptor expression and function in human skin and the pilosebaceous unit. *Exp Dermatol* 16:844–852
 110. Schneider MR, Paus R (2010) Sebocytes, multifaceted epithelial cells: lipid production and holocrine secretion. *Int J Biochem Cell Biol* 42:181–185
 111. Schneider MR, Schmidt-Ullrich R, Paus R (2009) The hair follicle as a dynamic miniorgan. *Curr Biol* 19:R132–R142
 112. Schoonjans K, Staels B, Auwerx J (1996) The peroxisome proliferator activated receptors (PPARs) and their effects on lipid metabolism and adipocyte differentiation. *Biochim Biophys Acta* 1302:93–109
 113. Seiffert K, Seltmann H, Fritsch M, Zouboulis CC (2007) Inhibition of 5alpha-reductase activity in SZ95 sebocytes and HaCaT keratinocytes in vitro. *Horm Metab Res* 39:141–148
 114. Seltmann H, Menon GK, Zouboulis CC (2005) Ca²⁺ regulates SZ95 sebocyte numbers and differentiation in an inverse manner compared to human keratinocytes. *J Invest Dermatol* 124:A108–A108
 115. Sertznig P, Seifert M, Tilgen W, Reichrath J (2009) Activation of vitamin D receptor (VDR)- and peroxisome proliferator-activated receptor (PPAR)-signaling pathways through 1, 25(OH)(2)D(3) in melanoma cell lines and other skin-derived cell lines. *Dermatoendocrinol* 1:232–238
 116. Slominski A (2005) Neuroendocrine system of the skin. *Dermatology* 211:199–208
 117. Slominski A (2009) On the role of the corticotropin-releasing hormone signalling system in the aetiology of inflammatory skin disorders. *Br J Dermatol* 160:229–232
 118. Slominski A, Ermak G, Hwang J, Chakraborty A, Mazurkiewicz JE, Mihm M (1995) Proopiomelanocortin, corticotropin releasing hormone and corticotropin releasing hormone receptor genes are expressed in human skin. *FEBS Lett* 374:113–116
 119. Slominski A, Ermak G, Hwang J, Mazurkiewicz J, Corliss D, Eastman A (1996) The expression of proopiomelanocortin (POMC) and of corticotropin releasing hormone receptor (CRH-R) genes in mouse skin. *Biochim Biophys Acta* 1289:247–251
 120. Slominski A, Ermak G, Mazurkiewicz JE, Baker J, Wortsman J (1998) Characterization of corticotropin-releasing hormone (CRH) in human skin. *J Clin Endocrinol Metab* 83:1020–1024
 121. Slominski A, Ermak G, Mihm M (1996) ACTH receptor, CYP11A1, CYP17 and CYP21A2 genes are expressed in skin. *J Clin Endocrinol Metab* 81:2746–2749
 122. Slominski A, Gomez-Sanchez CE, Foecking MF, Wortsman J (2000) Active steroidogenesis in the normal rat skin. *Biochim Biophys Acta* 1474:1–4
 123. Slominski A, Malarkey WB, Wortsman J, Asa SL, Carlson A (2000) Human skin expresses growth hormone but not the prolactin gene. *J Lab Clin Med* 136:476–481
 124. Slominski A, Paus R, Mazurkiewicz J (1992) Proopiomelanocortin expression in the skin during induced hair growth in mice. *Experientia* 48:50–54

125. Slominski A, Pisarchik A, Tobin DJ, Mazurkiewicz JE, Wortsman J (2004) Differential expression of a cutaneous corticotropin-releasing hormone system. *Endocrinology* 145:941–950
126. Slominski A, Roloff B, Curry J, Dahiya M, Szczesniwski A, Wortsman J (2000) The skin produces urocortin. *J Clin Endocrinol Metab* 85:815–823
127. Slominski A, Wortsman J (2000) Neuroendocrinology of the skin. *Endocr Rev* 21:457–487
128. Slominski A, Wortsman J, Luger T, Paus R, Solomon S (2000) Corticotropin releasing hormone and proopiomelanocortin involvement in the cutaneous response to stress. *Physiol Rev* 80:979–1020
129. Slominski A, Wortsman J, Paus R, Elias PM, Tobin DJ, Feingold KR (2008) Skin as an endocrine organ: implications for its function. *Drug Discov Today Dis Mech* 5:137–144
130. Slominski A, Wortsman J, Pisarchik A, Zbytek B, Linton EA, Mazurkiewicz JE, Wei ET (2001) Cutaneous expression of corticotropin-releasing hormone (CRH), urocortin, and CRH receptors. *FASEB J* 15:1678–1693
131. Slominski A, Wortsman J, Tuckey RC, Paus R (2007) Differential expression of HPA axis homolog in the skin. *Mol Cell Endocrinol* 265–266:143–149
132. Slominski A, Zbytek B, Semak I, Sweatman T, Wortsman J (2005) CRH stimulates POMC activity and corticosterone production in dermal fibroblasts. *J Neuroimmunol* 162:97–102
133. Slominski A, Zbytek B, Szczesniwski A, Semak I, Kaminski J, Sweatman T, Wortsman J (2005) CRH stimulation of corticosteroids production in melanocytes is mediated by ACTH. *Am J Physiol Endocrinol Metab* 288:E701–E706
134. Slominski A, Zbytek B, Szczesniwski A, Wortsman J (2006) Cultured human dermal fibroblasts do produce cortisol. *J Invest Dermatol* 126:1177–1178
135. Slominski A, Zjawiony J, Wortsman J, Semak I, Stewart J, Pisarchik A, Sweatman T, Marcos J, Dunbar C, Tuckey R (2004) A novel pathway for sequential transformation of 7-dehydrocholesterol and expression of the P450scc system in mammalian skin. *Eur J Biochem* 271:4178–4188
136. Smith KR, Thiboutot DM (2008) Thematic review series: skin lipids. Sebaceous gland lipids: friend or foe? *J Lipid Res* 49:271–281
137. Smith TM, Cong Z, Gilliland KL, Clawson GA, Thiboutot DM (2006) Insulin-like growth factor-1 induces lipid production in human SEB-1 sebocytes via sterol response element-binding protein-1. *J Invest Dermatol* 126:1226–1232
138. Smith TM, Gilliland K, Clawson GA, Thiboutot D (2008) IGF-1 induces SREBP-1 expression and lipogenesis in SEB-1 sebocytes via activation of the phosphoinositide 3-kinase/Akt pathway. *J Invest Dermatol* 128:1286–1293
139. Soberman RJ, Christmas P (2003) The organization and consequences of eicosanoid signaling. *J Clin Investig* 111:1107–1113
140. Stander S, Bohm M, Brzoska T, Zimmer KP, Luger T, Metze D (2002) Expression of melanocortin-1 receptor in normal, malformed and neoplastic skin glands and hair follicles. *Exp Dermatol* 11:42–51
141. Stander S, Moormann C, Schumacher M, Buddenkotte J, Artuc M, Shpacovitch V, Brzoska T, Lippert U, Henz BM, Luger TA, Metze D, Steinhoff M (2004) Expression of vanilloid receptor subtype 1 in cutaneous sensory nerve fibers, mast cells, and epithelial cells of appendage structures. *Exp Dermatol* 13:129–139
142. Stander S, Schmelz M, Metze D, Luger T, Rukwied R (2005) Distribution of cannabinoid receptor 1 (CB1) and 2 (CB2) on sensory nerve fibers and adnexal structures in human skin. *J Dermatol Sci* 38:177–188
143. Telek A, Biro T, Bodo E, Toth BI, Borbiro I, Kunos G, Paus R (2007) Inhibition of human hair follicle growth by endo- and exocannabinoids. *FASEB J* 21:3534–3541
144. Thiboutot D, Jabara S, McAllister JM, Sivarajah A, Gilliland K, Cong Z, Clawson G (2003) Human skin is a steroidogenic tissue: steroidogenic enzymes and cofactors are expressed in epidermis, normal sebocytes, and an immortalized sebocyte cell line (SEB-1). *J Invest Dermatol* 120:905–914
145. Thiboutot D, Sivarajah A, Gilliland K, Cong Z, Clawson G (2000) The melanocortin 5 receptor is expressed in human sebaceous glands and rat preputial cells. *J Invest Dermatol* 115:614–619
146. Thiele JJ, Weber SU, Packer L (1999) Sebaceous gland secretion is a major physiologic route of vitamin E delivery to skin. *J Invest Dermatol* 113:1006–1010
147. Thody AJ, Shuster S (1989) Control and function of sebaceous glands. *Physiol Rev* 69:383–416
148. Tomkova H, Kohoutek M, Zabojsnikova M, Pospiskova M, Ostrizkova L, Gharibayr M (2010) Cetuximab-induced cutaneous toxicity. *J Eur Acad Dermatol Venereol* 24:692–696
149. Toth BI, Geczy T, Griger Z, Dozsa A, Seltmann H, Kovacs L, Nagy L, Zouboulis CC, Paus R, Biro T (2009) Transient receptor potential vanilloid-1 signaling as a regulator of human sebocyte biology. *J Invest Dermatol* 129:329–339
150. Toyoda M, Morohashi M (2001) Pathogenesis of acne. *Med Electron Microsc* 34:29–40
151. Trivedi NR, Cong Z, Nelson AM, Albert AJ, Rosamilia LL, Sivarajah S, Gilliland KL, Liu W, Mauger DT, Gabbay RA, Thiboutot DM (2006) Peroxisome proliferator-activated receptors increase human sebum production. *J Invest Dermatol* 126:2002–2009
152. van Bilsen M, van der Vusse GJ, Gilde AJ, Lindhout M, van der Lee KAJM (2002) Peroxisome proliferator-activated receptors: lipid binding proteins controlling gene expression. *Mol Cell Biochem* 239:131–138
153. Wheatley VR, Potter JE, Lew G (1979) Sebaceous gland differentiation: II. The isolation, separation and characterization of cells from the mouse preputial gland. *J Invest Dermatol* 73:291–296
154. Wille JJ, Kydonieus A (2003) Palmitoleic acid isomer (C16:1delta6) in human skin sebum is effective against gram-positive bacteria. *Skin Pharmacol Appl Skin Physiol* 16:176–187
155. Wintzen M, Gilchrist BA (1996) Proopiomelanocortin, its derived peptides, and the skin. *J Invest Dermatol* 106:3–10
156. Wrobel A, Seltmann H, Fimmel S, Muller-Decker K, Tsukada M, Bogdanoff B, Mandt N, Blume-Peytavi U, Orfanos CE, Zouboulis CC (2003) Differentiation and apoptosis in human immortalized sebocytes. *J Invest Dermatol* 120:175–181
157. Yuspa SH, Hennings H, Tucker RW, Jaken S, Kilkenny AE, Roop DR (1988) Signal transduction for proliferation and differentiation in keratinocytes. *Ann N Y Acad Sci* 548:191–196
158. Zhang L, Anthonavage M, Huang Q, Li WH, Eisinger M (2003) Proopiomelanocortin peptides and sebogenesis. *Ann N Y Acad Sci* 994:154–161
159. Zhang L, Li WH, Anthonavage M, Eisinger M (2006) Melanocortin-5 receptor: a marker of human sebocyte differentiation. *Peptides* 27:413–420
160. Zhang Q, Seltmann H, Zouboulis CC, Konger RL (2006) Involvement of PPARgamma in oxidative stress-mediated prostaglandin E(2) production in SZ95 human sebaceous gland cells. *J Invest Dermatol* 126:42–48
161. Zouboulis CC (2004) Acne and sebaceous gland function. *Clin Dermatol* 22:360–366
162. Zouboulis CC (2009) Sebaceous gland receptors. *Dermatoendocrinol* 1:77–80
163. Zouboulis CC (2009) The skin as an endocrine organ. *Dermatoendocrinol* 1:250–252

164. Zouboulis CC, Akamatsu H, Stephanek K, Orfanos CE (1994) Androgens affect the activity of human sebocytes in culture in a manner dependent on the localization of the sebaceous glands and their effect is antagonized by spironolactone. *Skin Pharmacol* 7:33–40
165. Zouboulis CC, Baron JM, Bohm M, Kippenberger S, Kurzen H, Reichrath J, Thielitz A (2008) Frontiers in sebaceous gland biology and pathology. *Exp Dermatol* 17:542–551
166. Zouboulis CC, Bohm M (2004) Neuroendocrine regulation of sebocytes—a pathogenetic link between stress and acne. *Exp Dermatol* 13(Suppl 4):31–35
167. Zouboulis CC, Chen WC, Thornton MJ, Qin K, Rosenfield R (2007) Sexual hormones in human skin. *Horm Metab Res* 39:85–95
168. Zouboulis CC, Eady A, Philpott M, Goldsmith LA, Orfanos C, Cunliffe WC, Rosenfield R (2005) What is the pathogenesis of acne? *Exp Dermatol* 14:143–152
169. Zouboulis CC, Schagen S, Aletas T (2008) The sebocyte culture: a model to study the pathophysiology of the sebaceous gland in seborrhoea, seborrhoea and acne. *Arch Dermatol Res* 300:397–413
170. Zouboulis CC, Seltmann H, Hiroi N, Chen W, Young M, Oeff M, Scherbaum WA, Orfanos CE, McCann SM, Bornstein SR (2002) Corticotropin-releasing hormone: an autocrine hormone that promotes lipogenesis in human sebocytes. *Proc Natl Acad Sci U S A* 99:7148–7153
171. Zouboulis CC, Seltmann H, Neitzel H, Orfanos CE (1999) Establishment and characterization of an immortalized human sebaceous gland cell line (SZ95). *J Invest Dermatol* 113:1011–1020
172. Zouboulis CC, Xia L, Akamatsu H, Seltmann H, Fritsch M, Hornemann S, Ruhl R, Chen W, Nau H, Orfanos CE (1998) The human sebocyte culture model provides new insights into development and management of seborrhoea and acne. *Dermatology* 196:21–31
173. Zygmunt PM, Petersson J, Andersson DA, Chuang H, Sorgard M, Di Marzo V, Julius D, Hogestatt ED (1999) Vanilloid receptors on sensory nerves mediate the vasodilator action of anandamide. *Nature* 400:452–45